

Synthesis of Cyclic Aminomethylphosphonates and Aminomethyl-Arylphosphinic Acids by an Efficient Microwave-Mediated Phospha-Mannich Approach

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ABSTRACT: Microwave-assisted condensation of 1,3,2-dioxaphosphinane 2-oxide (**1**), paraformaldehyde and secondary amines including 5- and 6-membered *N*-heterocycles at 55°C gave cyclic aminomethylphosphonates (**2**), whereas an analogous reaction involving dibenzo[*c.e*][1,2]oxaphosphinane 2-oxide (**3**) resulted in the corresponding aminomethyl-2-(2'-hydroxybiphenyl)phosphinic acids (**4**) as a consequence of a hydrolytic ring opening following the condensation. © 2008 Wiley Periodicals, Inc. *Heteroatom Chem* 19:207–210, 2008; Published online in Wiley InterScience (www.interscience.wiley.com). DOI 10.1002/hc.20387

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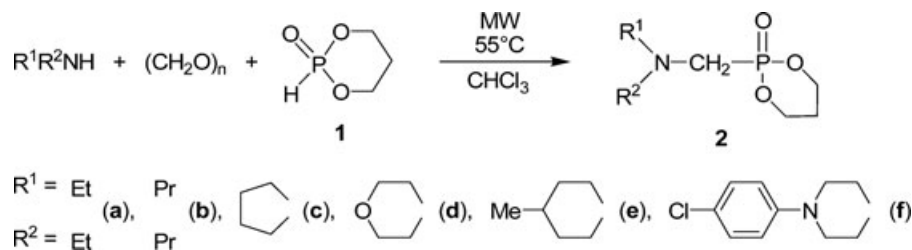
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INTRODUCTION

The Kabachnik–Fields reaction [1,2] involving the condensation of a ternary system comprising a $>P(O)H$ reactant, an aldehyde or ketone, and a secondary or primary amine is an important method for the synthesis of compounds of type of $>NC(R^1)(R^2)P(O)<$, such as aminophosphonates [3–6] that forms an important class of biologically active compounds. The green chemical aspects of the Kabachnik–Fields reaction also received considerable attention [4,6].

In our laboratory, the microwave promoted, solvent-free synthesis of phosphono- and phosphinoylmethylated *N*-heterocycles has been elaborated [7]. In this paper, we introduce aminomethylphosphonic and phosphinic derivatives based or related on 6-membered P-heterocycles.

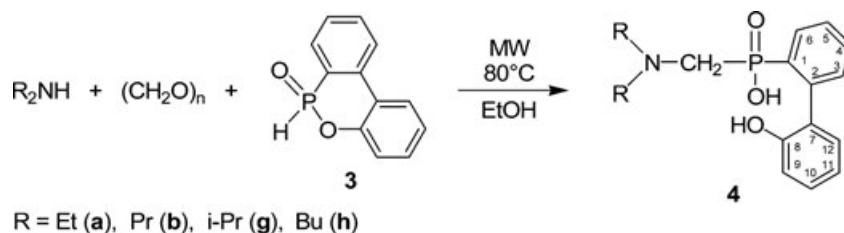
On the basis of our previous experiences with cyclic $>P(O)H$ species [8,9], it was a challenge for us to convert them to the corresponding aminomethyl derivatives. 1,3,2-Dioxaphosphinane



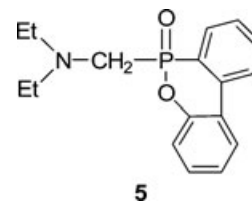
SCHEME 1

2-oxide **1** reacted easily with paraformaldehyde and secondary amines including pyrrolidine, morpholine, piperidine-, and piperazine derivatives at 55°C in chloroform under microwave to afford the corresponding aminomethylphosphonates **2a–f** (Scheme 1). After purification by column chromatography, products **2a–f** were obtained in 54%–72% yields and were characterized by ^{31}P , ^{13}C , and 1H NMR, as well as mass spectral data.

The next cyclic $>P(O)H$ species was dibenzo[*c.e*][1,2]oxaphosphinane oxide **3** that was reacted with paraformaldehyde and secondary amines at 80°C in dry ethanol, again under microwave. Instead of the desired aminomethyl-dibenzooxaphosphorine oxides, aminomethyl-2-(2'-hydroxybiphenyl)phosphinic acids **4a,b,g,h** were obtained (Scheme 2). Compounds **4** were formed by opening of the oxaphosphinane ring of the primary product (e.g., **5**) via hydrolysis. The water bringing about the hydrolytic ring opening is derived from the primary Mannich condensation. Products **4a,b,g,h** were purified by recrystallization and were identified on the basis of ^{31}P , ^{13}C , and 1H NMR, as well as mass spectrometry. Carrying out the condensation in chloroform instead of ethanol, the ring opening was somewhat suppressed and the presence of the aminomethyldibenzooxaphosphinane oxide (e.g., **5**) could be detected by ^{31}P NMR and FAB, but it was not possible to prevent the predominant ring opening. The ratio of **4a** and **5** was ca. 75:25 on the basis of relative ^{31}P NMR intensities.



SCHEME 2



In summary, new aminomethyl cyclic phosphonates and hydroxybiphenylphosphinic acids were synthesized by the microwave-assisted condensation of a cyclic or acyclic secondary amine, paraformaldehyde, and the $P(O)H$ derivative of the corresponding P-heterocycle.

EXPERIMENTAL

The ^{31}P , ^{13}C , and 1H NMR spectra were obtained on a Bruker DRX-500 spectrometer operating at 202.4, 125.7, and 500 MHz, respectively. Chemical shifts are downfield relative to 85% H_3PO_4 or TMS. Mass spectrometry was performed on a ZAB-2SEQ instrument. The reactions were carried out in a 300-W CEM Discover focused microwave reactor under isothermic conditions.

General Procedure for the Preparation of Aminomethyl[1,3,2]dioxaphosphinane 2-Oxides (**2a–f**)

A mixture of [1,3,2]dioxaphosphinane 2-oxide **1** [10] (0.21 g, 1.7 mmol), paraformaldehyde (0.05 g,

1.7 mmol), and secondary amine (1.7 mmol) in 2 mL of chloroform was stirred at 55°C in a microwave reactor for 20 min. Volatile components including water were removed in vacuo. Column chromatography (silica gel, 3% methanol in chloroform) of the residue afforded the product (**2a–f**) as an oil.

The following products were thus prepared:

2a: Yield, 72%; ^{31}P NMR (CDCl_3) δ 21.5; ^{13}C NMR (CDCl_3) δ 11.6 (CH_3), 26.6 ($^3J = 8.4$, $\text{CH}_2\text{CH}_2\text{CH}_2$), 48.6 ($^3J = 8.8$, NCH_2CH_3), 49.0 ($^1J = 159.7$, PCH_2), 67.2 ($^2J = 7.5$, OCH_2); ^1H NMR (CDCl_3) δ 1.08 (t, $J = 7.0$, 6H, CH_3), 2.06–2.15 (m, 2H, $\text{CH}_2\text{CH}_2\text{CH}_2$), 2.69–2.78 (m, 4H, NCH_2CH_3), 3.03 (br d, $^2J_{\text{PH}} = 10.0$, 2H, PCH_2), 4.33–4.46 (m, 2H, OCH_2), 4.46–4.58 (m, 2H, OCH_2); ($\text{M} + \text{H}$) $^+_{\text{found}} = 208.1084$, $\text{C}_8\text{H}_{19}\text{NO}_3\text{P}$ requires 208.1102.

2b: Yield: 55%; ^{31}P NMR (CDCl_3) δ 19.6; ^{13}C NMR (CDCl_3) δ 11.3 (CH_3), 19.7 ($\text{CH}_2\text{CH}_2\text{CH}_3$), 26.2 ($^3J = 8.2$, $\text{CH}_2\text{CH}_2\text{CH}_2$), 49.9 ($^1J = 157.9$, PCH_2), 57.0 ($^3J = 8.0$, NCH_2CH_2), 66.3 ($^2J = 7.3$, OCH_2); ^1H NMR (CDCl_3) δ 0.90 (t, $J = 7.5$, 6H, CH_3), 1.40–1.55 (m, 4H, $\text{CH}_2\text{CH}_2\text{CH}_3$), 2.00–2.18 (m, 2H, $\text{CH}_2\text{CH}_2\text{CH}_2$), 2.54 (m, 4H, NCH_2CH_2), 2.99 (d, $^2J_{\text{PH}} = 9.9$, 2H, PCH_2), 4.30–4.42 (m, 2H, OCH_2), 4.45–4.58 (m, 2H, OCH_2); ($\text{M} + \text{H}$) $^+_{\text{found}} = 236.1398$, $\text{C}_{10}\text{H}_{23}\text{NO}_3\text{P}$ requires 236.1416.

2c: Yield: 49%; ^{31}P NMR (CDCl_3) δ 20.9; ^{13}C NMR (CDCl_3) δ 23.9 (NCH_2CH_2), 26.7 ($^3J = 7.9$, $\text{CH}_2\text{CH}_2\text{CH}_2$), 51.2 ($^1J = 161.8$, PCH_2), 56.2 ($^3J = 10.7$, NCH_2CH_2), 66.6 ($^2J = 7.0$, OCH_2); ^1H (CDCl_3) NMR δ 1.75–1.85 (m, 4H, NCH_2CH_2), 2.00–2.16 (m, 2H, $\text{CH}_2\text{CH}_2\text{CH}_2$), 2.65–2.75 (m, 4H, NCH_2CH_2), 3.05 (d, $^2J_{\text{PH}} = 12.0$, 2H, PCH_2), 4.30–4.42 (m, 2H, OCH_2), 4.49–4.61 (m, 2H, OCH_2); ($\text{M} + \text{H}$) $^+_{\text{found}} = 206.0937$, $\text{C}_8\text{H}_{17}\text{NO}_3\text{P}$ requires 206.0946.

2d: Yield: 50%; ^{31}P NMR (CDCl_3) δ 19.8; ^{13}C NMR (CDCl_3) δ 26.8 ($^3J = 8.1$, $\text{CH}_2\text{CH}_2\text{CH}_2$), 54.3 ($^1J = 160.6$, PCH_2), 55.4 ($^3J = 10.1$, $\text{OCH}_2\text{CH}_2\text{N}$), 66.8 ($^2J = 7.1$, POCH_2), 67.1 ($\text{OCH}_2\text{CH}_2\text{N}$); ^1H NMR (CDCl_3) δ 2.03–2.15 (m, 2H, $\text{CH}_2\text{CH}_2\text{CH}_2$), 2.65 (t, $^3J_{\text{HH}} = 4.8$, 4H, $\text{OCH}_2\text{CH}_2\text{N}$), 2.89 (d, $^2J_{\text{PH}} = 11.4$, 2H, PCH_2), 3.71 (t, $J_{\text{HH}} = 4.5$, 4H, $\text{OCH}_2\text{CH}_2\text{N}$), 4.29–4.43 (m, 2H, OCH_2), 4.47–4.61 (m, 2H, OCH_2); ($\text{M} + \text{H}$) $^+_{\text{found}} = 222.0880$, $\text{C}_8\text{H}_{17}\text{NO}_4\text{P}$ requires 222.0895.

2e: Yield: 54%; ^{31}P NMR (CDCl_3) δ 19.8; ^{13}C NMR (CDCl_3) δ 21.7 (CH_3), 26.5 ($^3J = 8.1$, $\text{CH}_2\text{CH}_2\text{CH}_2$), 30.0 (CH), 34.4 (CHCH_2), 54.3 ($^1J = 159.6$, PCH_2), 55.7 ($^3J = 9.6$, NCH_2), 67.3 ($^2J = 7.3$, OCH_2); ^1H NMR (CDCl_3) δ 0.90 (d, $J = 6.4$, 3H, CH_3), 1.18–1.28 and 1.58–1.64 (m, 4H, CHCH_2), 1.29–1.40 (m, 1H, CH), 2.02–2.16 (m, 2H, $\text{CH}_2\text{CH}_2\text{CH}_2$), 2.21 (br t, $J = 11.6$, 2H, $\text{CH}_2\text{CH}_2\text{N}$), 2.89 (d, $^1J_{\text{PH}} = 10.2$, 2H, PCH_2), 2.97 (br d, $J = 11.5$, 2H, $\text{CH}_2\text{CH}_2\text{N}$), 4.38–4.56

(m, 4H, OCH_2); ($\text{M} + \text{H}$) $^+_{\text{found}} = 234.1245$, $\text{C}_{10}\text{H}_{21}\text{NO}_3\text{P}$ requires 234.1259.

2f: Yield: 64%; ^{31}P NMR (CDCl_3) δ 19.1; ^{13}C NMR (CDCl_3) δ 26.5 ($^3J = 8.3$, $\text{CH}_2\text{CH}_2\text{CH}_2$), 49.1 (ArNCH_2), 52.6 ($^1J = 159.7$, PCH_2), 54.7 ($^3J = 10.0$, $\text{CH}_2\text{NCH}_2\text{P}$), 67.0 ($^2J = 7.2$, OCH_2), 117.3 (CHCN), 124.5 (ClC), 128.9 (ClCCH), 149.7 (NC); ^1H NMR (CDCl_3) δ 2.10–2.10 (m, 2H, $\text{CH}_2\text{CH}_2\text{CH}_2$), 2.65 (t, $^3J_{\text{HH}} = 5.0$, 4H, $\text{CH}_2\text{NCH}_2\text{P}$), 2.97 (d, $^2J_{\text{PH}} = 11.5$, 2H, PCH_2), 3.19 (t, $^3J_{\text{HH}} = 5.5$, 4H, $\text{ArNCH}_2\text{CH}_2\text{N}$), 4.35–4.44 (m, 2H, OCH_2), 4.47–4.59 (m, 2H, OCH_2), 6.82 (d, $J_{\text{HH}} = 9.0$, 2H, Ar), 7.20 (d, $J_{\text{HH}} = 9.0$, 2H, Ar); ($\text{M} + \text{H}$) $^+_{\text{found}} = 331.0956$, $\text{C}_{14}\text{H}_{21}\text{N}_2\text{O}_3\text{P}$ requires 331.0978.

General Procedure for the Preparation of Aminomethyl-2-(2'-hydroxybiphenyl)phosphinic Acids **4a,b,g,h**

A mixture of dibenzooxaphosphinane oxide **3** (0.20 g, 0.93 mmol), paraformaldehyde (0.03 g, 1.0 mmol), and secondary amine (1.0 mmol) in 2 mL of ethanol was reacted under microwave conditions at 80°C for 1.5 h. Volatile components including water were removed in vacuo to give phosphinic acid (**4a,b,g,h**) as a solid.

The following products were thus prepared:

4a: Yield, 98%; mp. 176°C–178°C after recrystallization from ethyl acetate–pentane; ^{31}P NMR (CDCl_3) δ 15.1; ^{13}C NMR (CDCl_3) δ 7.8 (CH_3), 47.9 ($^3J = 4.4$, NCH_2CH_3), 50.8 ($^1J = 91.3$, PCH_2), 121.2 (Ar),^a 122.5 (Ar),^a 127.2 ($J = 11.3$, Ar),^b 129.6 (Ar),^a 130.9 (Ar),^a 131.2 ($J = 2.4$, Ar),^b 131.7 ($J = 7.6$, Ar),^b 132.1 ($J = 11.1$, Ar),^b 134.2 ($^2J = 3.0$, C_2), 135.5 ($^1J = 129.2$, C_1), 141.9 ($^3J = 10.3$, C_7), 154.9 (C_8),^a tentative assignment for C_9 , C_{10} , C_{11} and C_{12} ,^b tentative assignment for C_3 , C_4 , C_5 , and C_6 ; ^1H NMR (CDCl_3) δ 0.97–1.15 (m, 6H, CH_3), 2.66–2.76 (m, 2H, PCH_2), 2.93–3.18 (m, 4H, NCH_2CH_3), 7.04 (t, $J = 7.5$, 1H, ArH), 7.11 (d, $J = 6.5$, 1H, ArH), 7.14 (d, $J = 8.5$, 1H, ArH), 7.23 (dd, $J_1 = 7.0$, $J_2 = 5.0$, 1H, ArH), 7.33 (t, $J = 7.0$, 1H, ArH), 7.43 (t, $J = 7.5$, 1H, ArH), 7.51 (t, $J = 7.5$, 1H, ArH), 8.08 (dd, $J_1 = 7.5$, $J_2 = 12.5$, 1H, ArH); ($\text{M} + \text{H}$) $^+_{\text{found}} = 320.1402$, $\text{C}_{17}\text{H}_{23}\text{NO}_3\text{P}$ requires 320.1416. Performing the above reaction at 55°C for 40 min in 2 mL of chloroform led to a mixture of **4a** (75%) and **5** (25%).

5: ^{31}P NMR (CDCl_3) δ 33.8; ($\text{M} + \text{H}$) $^+_{\text{found}} = 302.1278$, $\text{C}_{17}\text{H}_{21}\text{NO}_2\text{P}$ requires 302.1310.

4b: Yield, 56%; mp 156°C–158°C after crystallization from ethylacetate–hexane; ^{31}P NMR (CDCl_3) δ 14.9; ^{13}C NMR (CDCl_3) δ 10.7 (CH_3), 16.1 (CH_2CH_2), 52.0 ($^1J = 90.8$, PCH_2), 55.6 (br signal, NCH_2CH_2), 121.5 (Ar),^a 121.7 (Ar),^a 127.4 ($J = 11.3$, Ar),^b 129.7 (Ar),^a 131.1 (Ar),^a 131.5 ($J = 1.8$, Ar),^b 131.9 ($J = 8.0$,

Ar),^b 132.2 ($J = 11.5$, Ar),^b 133.7 ($^2J = 3.0$, C₂), 134.9 ($^1J = 130.7$, C₁), 141.6 ($^3J = 10.5$, C₇), 154.2 (C₈),^atentative assignment for C₉, C₁₀, C₁₁, and C₁₂,^btentative assignment for C₃, C₄, C₅, and C₆; ¹H NMR (CDCl₃) δ 0.68–0.85 (m, 6H, CH₃), 1.37–1.58 (m, 4H, CH₂CH₃), 2.67–2.98 (overlapping signals, 6H, NCH₂ + PCH₂), 6.98–7.52 (m, 8H, ArH), 8.05 (dd, $J_1 = 7.5$, $J_2 = 11.5$, 1H, OH), 10.40 (br s, 1H, OH); (M + H)⁺_{found} = 348.1709, C₁₉H₂₇NO₃P requires 348.1729.

4g: Yield: 59%; ³¹P NMR (CDCl₃) δ 13.3; ¹³C NMR (CDCl₃) δ 17.8 (br s, CH₃), 46.0 ($^1J = 87.3$, PCH₂), 55.7 (NCH), 121.2 (Ar),^a 122.2 (Ar),^a 127.1 ($J = 11.5$, Ar),^b 129.6 (Ar),^a 130.8 (Ar),^a 131.0 ($J = 2.3$, Ar),^b 131.5 ($J = 7.7$, Ar),^b 131.8 ($J = 11.1$, Ar),^b 134.0 ($^2J = 3.0$, C₂), 135.3 ($^1J = 132.1$, C₁), 141.2 ($^3J = 10.3$, C₇), 154.2 (C₈),^atentative assignment for C₉, C₁₀, C₁₁, and C₁₂,^btentative assignment for C₃, C₄, C₅, and C₆; ¹H NMR (CDCl₃) δ 1.12–~ 1.20 (m, 6H, CH₃), 1.21 (d, $J = 6.6$, 6H, CH₃), 2.52–2.64 (septet, $J = 7.7$, 2H, CH), 3.54–3.64 (m, 2H, PCH₂), 6.98–7.50 (m, 8H, Ar), 8.03 (dd, $J_1 = 7.6$, $J_2 = 11.5$, 1H, OH), 10.43 (br s, 1H, OH); (M + H)⁺_{found} = 348.1724, C₁₉H₂₇NO₃P requires 348.1729.

4h: Yield: 61%; mp. 152°C–154°C; ³¹P NMR (CDCl₃) δ 14.5; ¹³C NMR (CDCl₃) δ 13.5 (CH₃), 19.8 (CH₂CH₂), 24.2 (CH₂CH₂), 51.4 ($^1J = 92.3$, PCH₂), 52.9 (br s, NCH₂CH₂), 121.1 (Ar),^a 122.8 (Ar),^a 127.0 ($J = 11.2$, Ar),^b 129.5 (Ar),^a 130.7 (Ar),^a 131.0 ($J = 2.4$, Ar),^b 131.7 ($J = 7.5$, Ar),^b 132.1 ($J = 11.1$, Ar),^b 134.3 ($^2J = 2.9$, C₂), 135.6 ($^1J = 128.4$, C₁), 141.9 ($^3J = 10.3$, C₇), 155.0 (C₈),^atentative assignment for C₉, C₁₀, C₁₁, and C₁₂,^btentative assignment for C₃, C₄, C₅, and C₆; ¹H NMR (CDCl₃) δ 0.84 (br s, 6H,

CH₃), 0.98–1.80 (m, 8H, CH₂), 2.71–3.08 (overlapping signals, 6H, NCH₂ + PCH₂), 6.98–7.52 (m, 8H, Ar), 8.03 (dd, $J_1 = 7.4$, $J_2 = 12.3$, 1H, OH), 10.3 (br s, 1H, OH); (M + H)⁺_{found} = 376.2018, C₂₁H₃₁NO₃P requires 376.2042.

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